

Whole Transcriptome Analysis QC Report

QC Report for RNA-Seq Libraries generated using Micro-dissected Tissue

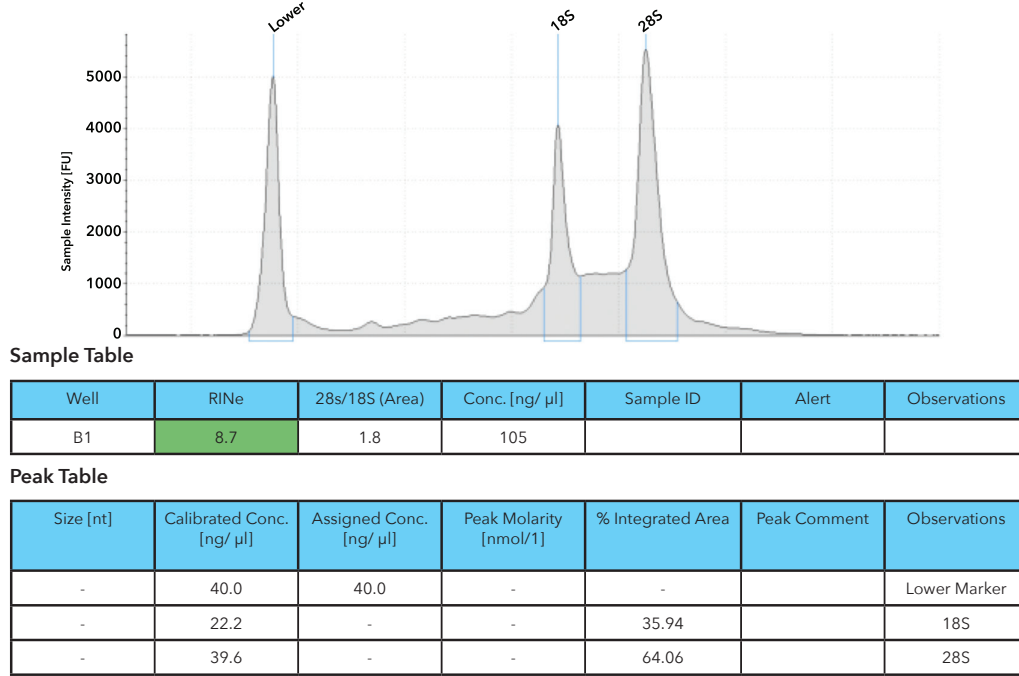


Figure 1: A) Representative Electropherogram generated from TapeStation shows profile of RNA input from micro-dissected tissue. B) Table shows quantification of the sample input and quality of RNA is reported by RIN. C) Table shows quantification of the peaks in the sample.

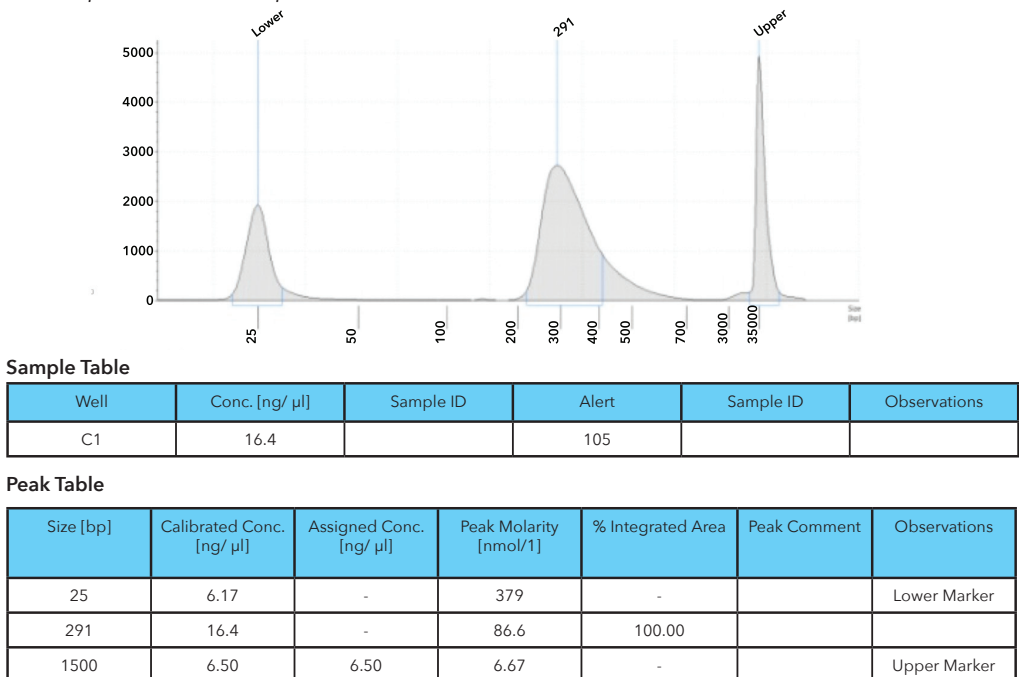
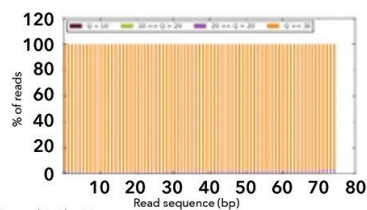


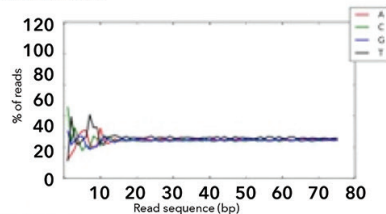
Figure 2: A) Representative Electropherogram generated from TapeStation shows profile of NGS library generated using the RNA isolated from fresh-frozen tissue. B) Table shows quantification of the library. C) Table shows the quantification of the peaks in the library.

8.1 Read orientation-R1

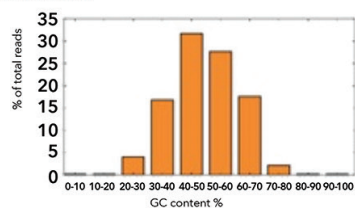
8.1.1 Quality distribution



8.1.2 Base distribution

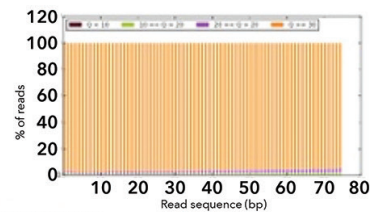


8.1.3 GC distribution

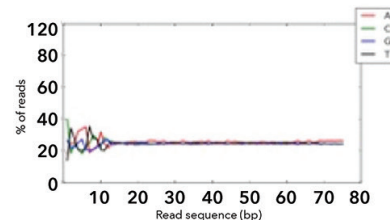


8.2 Read orientation-R2

8.2.1 Quality distribution



8.2.2 Base distribution



8.2.3 GC distribution

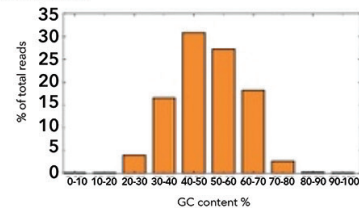
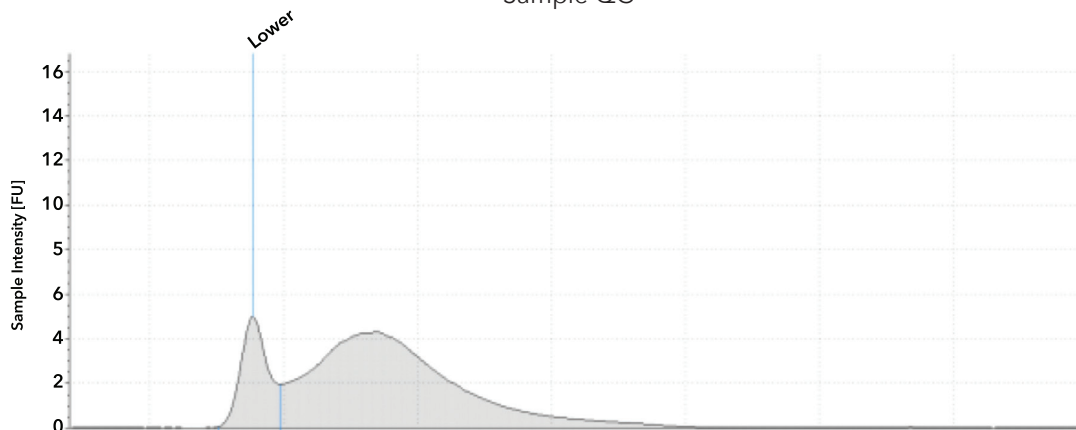


Figure 3: Representative FastQC report showing quality of reads obtained from the sequencing run.

QC Report for FFPE sample

Sample QC

A



B

Sample Table

Well	DV200	28s/18S (Area)	Conc. [ng/ μ l]	Sample ID	Alert	Observations
B1	58%	-	99.5			

C

Peak Table

Size [nt]	Calibrated Conc. [ng/ μ l]	Assigned Conc. [ng/ μ l]	Peak Molarity [nmol/l]	% Integrated Area	Peak Comment	Observations
-	40.0	40.0	-	-		Lower Marker

Figure 4: A) Representative Electropherogram generated from TapeStation shows profile of RNA input from FFPE-curl. B) Table shows quantification of the sample input and quality of RNA is reported by DV200. C) Table shows quantification of the peaks in the sample.

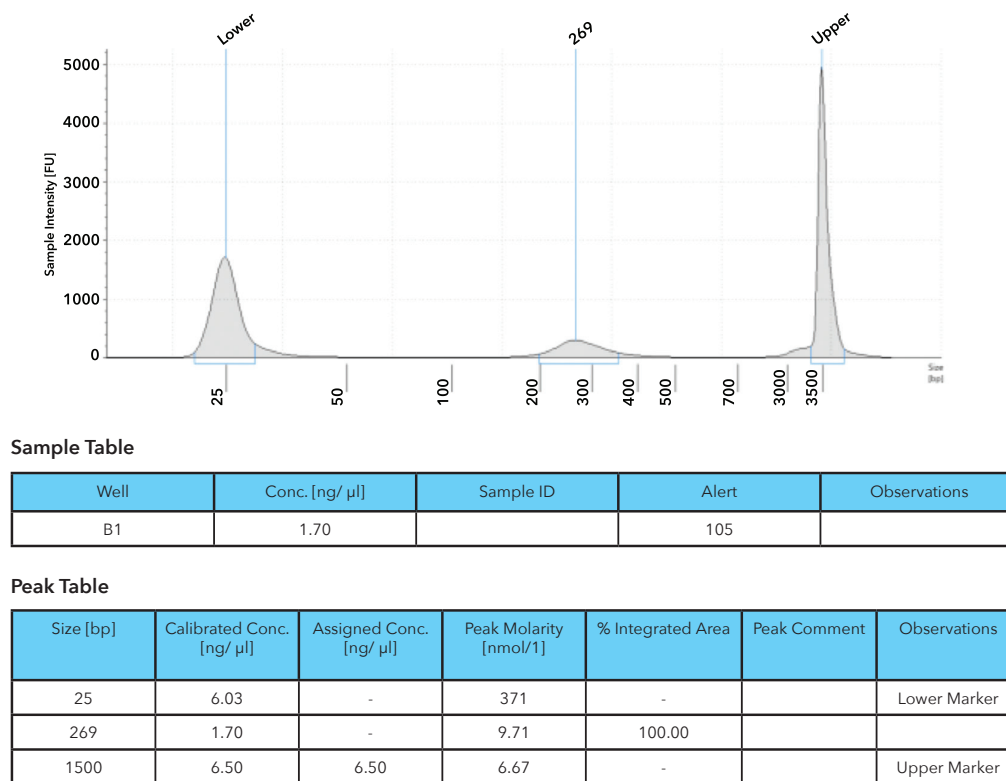
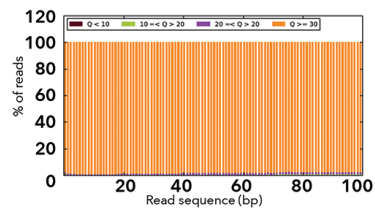


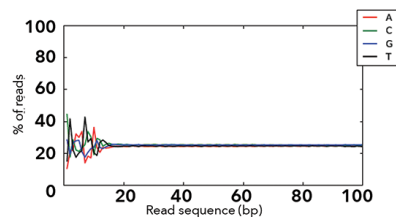
Figure 5: A) Representative Electropherogram generated from TapeStation shows profile of NGS library generated using the RNA isolated from fresh-frozen tissue. B) Table shows quantification of the library. C) Table shows the quantification of the peaks in the library.

2.1 Read orientation-R1

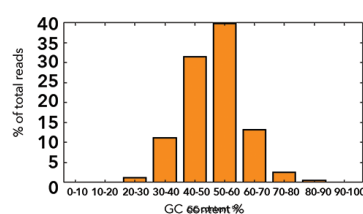
2.1.1 Quality distribution



2.1.2 Base distribution

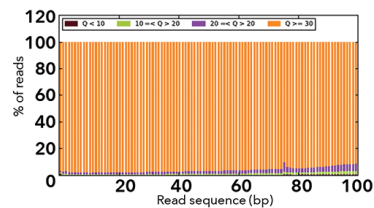


2.1.3 GC distribution

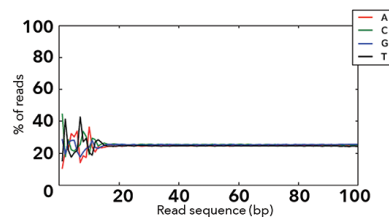


2.2 Read orientation-R2

2.2.1 Quality distribution



2.2.2 Base distribution



2.2.3 GC distribution

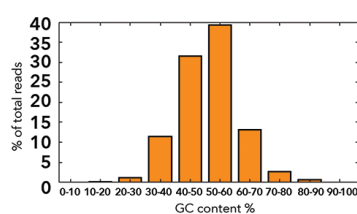


Figure 6: Representative FastQC report showing quality of reads obtained from the sequencing run.

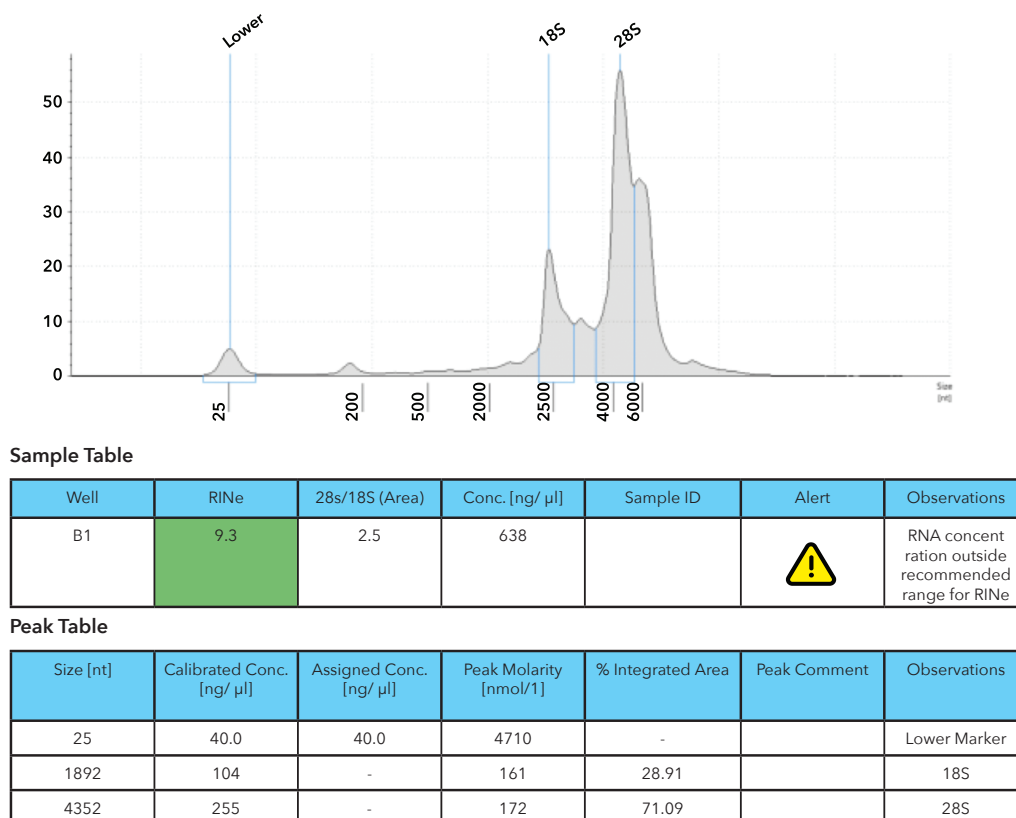


Figure 7: A) Representative Electrophoregram generated from TapeStation shows profile of RNA input from dissociated cells. B) Table shows quantification of the sample input and quality of RNA is reported by RIN. C) Table shows quantification of the peaks in the sample.

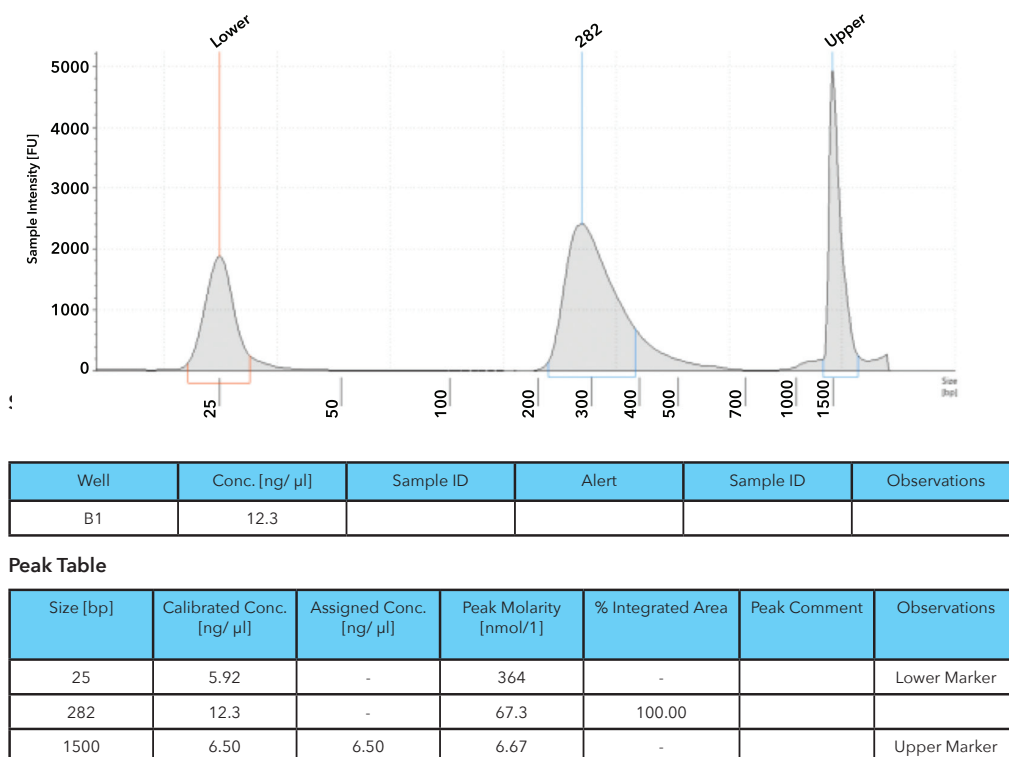
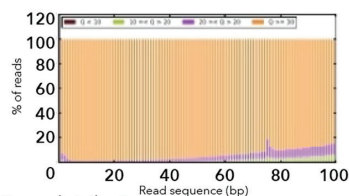


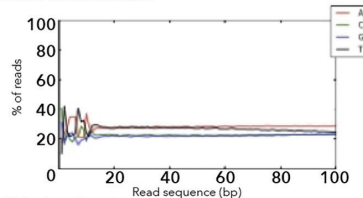
Figure 8: A) Representative Electrophoregram generated from TapeStation shows profile of NGS library generated using the RNA isolated from fresh-frozen tissue. B) Table shows quantification of the library. C) Table shows the quantification of the peaks in the library.

1.1 Read orientation-R1

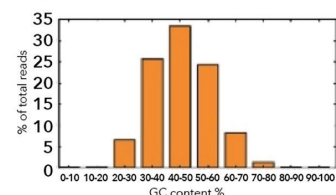
1.1.1 Quality distribution



1.1.2 Base distribution

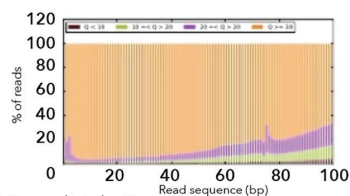


1.1.3 GC distribution

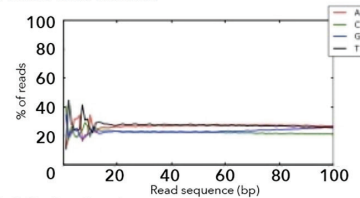


1.2 Read orientation-R2

1.2.1 Quality distribution



1.2.2 Base distribution



1.2.3 GC distribution

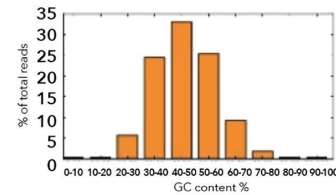
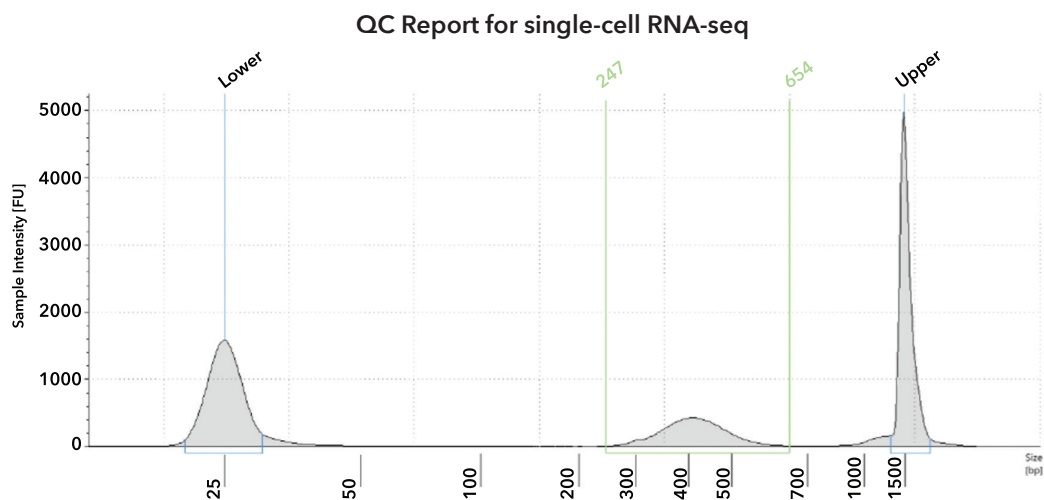


Figure 9: Representative FastQC report showing quality of reads obtained from the sequencing run.



Region Table

From [bp]	To [bp]	Average Size [bp]	Region Molarity [nmol/l]	Conc. [ng/μl]	% of Total	Region Comment	Color
247	654	417	3.22	12.2	73.74		

Figure 10: A) Representative electropherogram of gene expression libraries generated from single-cells using the 10X Gene Expression Analysis Kit. B) The table below shows quantification of the library.

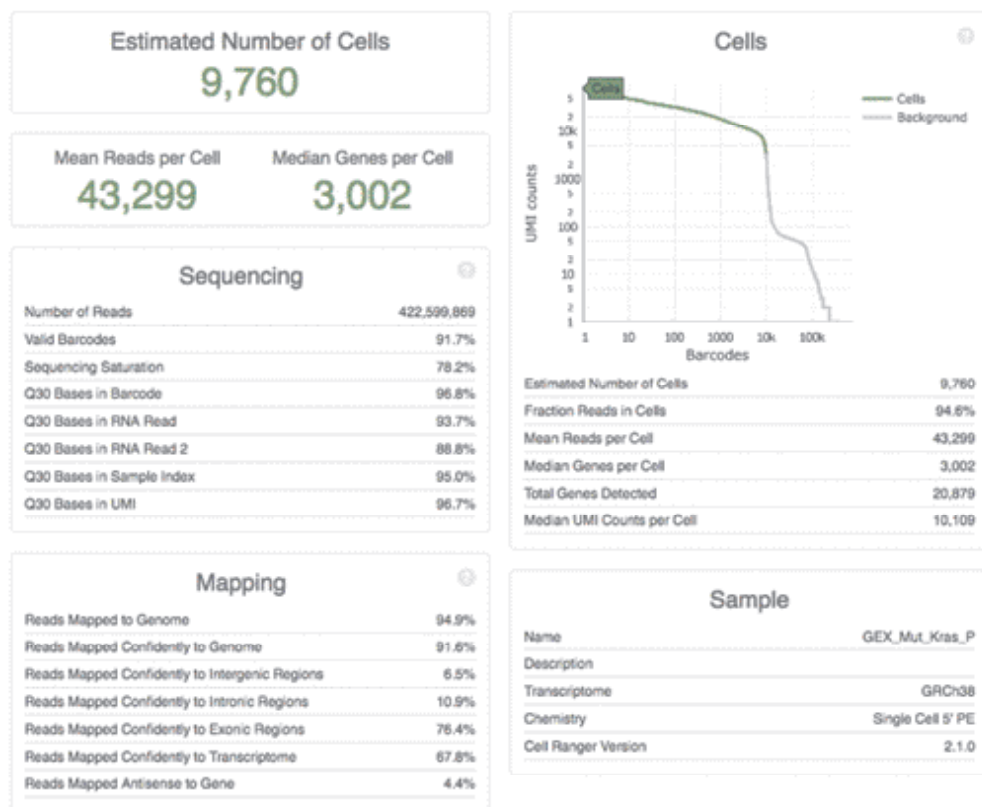


Figure 11: Sample representation of output from a 10X cell-ranger software of the summary of the reads obtained from a gene expression run with a 10X 3'-gene expression profiling kit.